

FLIPR FMP Blue Assay

1. Make 2 L Locke's buffer (with glycine for neurons). pH to 7.4.

Locke's buffer

| <u>Reagent</u> | <u>mM</u> | <u>1 Liter</u> | <u>2 Liters</u> |
|--|-----------|----------------|-----------------|
| HEPES | 8.6 | 2.144 g | 4.288 g |
| <u>KCl</u> | 5.6 | 0.417 g | 0.834 g |
| <u>NaCl</u> | 154 | 9.0 g | 18.0 g |
| Glucose | 5.6 | 1.009 g | 2.018 g |
| MgCl ₂ | 1.0 | 0.204 g | 0.408 g |
| CaCl ₂ | 2.3 | 0.338 g | 0.676 g |
| Glycine (primary neurons) | 0.1 | 7.5 mg | 15 mg |
| Probenecid (CHO Ca ²⁺ assays) | | 0.1175 g | 0.235 g |

2. Follow the steps on the machine for turning on and calibrating the machine.
3. To calibrate the machine, perform a yellow plate reading. First run the test without selecting calibration. Percent standard deviation will be high. Next run the calibration step. Percent standard deviation should be ~1.2%. Document max, average, min value and % standard deviation.
 - a. Filter1 for yellow plate reading and calcium oscillations
 - b. Filter2 for FMP blue.
4. Plan your experiment using 96-well plate template.
5. Create a file for each day experiments are performed. It is saved in Documents → Pierce → DATE
6. Select your experiment. If template is not made, will have to generate template.
7. To make FMP blue, add 1 FLIPR membrane potential assay kit to 20 mL Locke's buffer. Incubate 100 uL on cells for 45 minutes prior to experimentation.
8. Dilute out the drug(s).
 - a. For FMP blue, will do 100:20 uL
 - i. -5 dilution is 1.2 uL drug (50 mM stock) in 1 mL FMP dye.
 - ii. Serial dilute in 100 uL drug into 900 mL FMP dye.
9. Run on FLIPR2.
10. Analyze using GraphPad prism